

LSOP Title	Enzymatic Sugar Measurements
LSOP No.	LSOP09
Version	1.1
Location	UQ Node/Centre-wide
Policy/Procedure Link	<a href="#">UQ- Equipment</a> <a href="#">UQ -waste</a>
Risk Assessments	
Approved by	Franziska Fichtner
Date Approved	30/08/2021
Date Effective	03/06/2021
Next Review Date	03/06/2022
Contact for Assistance	<a href="mailto:plantsuccess@ug.edu.au">plantsuccess@ug.edu.au</a>

## 1.0 Scope


*This procedure covers the protocol for measuring enzymatic sugar concentrations*

*This CSOP does not cover the extraction process for samples or the use of the microplate reader.*

## 2.0 Definitions

G6PDH – Glucose-6-Phosphate Dehydrogenase

 HEPES - Sulfonic acid buffering agent

 ATP – Adenosine Tri-Phosphate

  KOH – Potassium Hydroxide

 MgCl<sub>2</sub> – Magnesium Chloride

NADP+ - Nicotinamide Adenine Dinucleotide Phosphate

NADPH – Reduced form of Nicotinamide Adenine Dinucleotide Phosphate

HXK – Hexokinase

PGI – Phosphoglucose Isomerase

INV – Invertase

### 3.0 Materials and Equipment

1. Microplate
2. Pipette & Pipette Tips
3. G6PDH in ammonium sulphate
4. Mix (see appendix for stocks)
5. Sugars (see appendix)
6. Microplate Reader
7. Centrifuge



### 4.0 Prescribed Actions

1. Prepare fresh: G6PDH grade II *from Roche in ammonium sulphate* [4°C]
  - a. To prepare the solution: centrifuge 2 min at 13,400 rpm, discard the supernatant and dissolve the pellet.
2. **Mix for 1 microplate:** 15.5 ml HEPES buffer, 480 µl ATP, 480 µl NADP, 80 µl G6PDH grade II (see appendix for stock solutions)
3. Pipette 150 µL of Mix (equivalent to 100 mM HEPES-KOH, pH 6.9, 3 mM MgCl<sub>2</sub>, 1 mM NADP<sup>+</sup>, 2.5 mM ATP, and 0.5 U glucose 6-phosphate dehydrogenase) into microtiter plate
4. Glucose, fructose and sucrose are determined by the sequential addition of
  - a. 0.9 U hexokinase (yeast)
  - b. 0.05 U phosphoglucose isomerase (yeast)
  - c. and 10 U invertase (yeast) to the amount of sugars.
5. Pipette 50 µL aliquots of sample extract into each well  
*NB: Generally dispensed as duplicates into a microtiter plate*
6. Mixed with the reaction mixture, the reduction of NADP<sup>+</sup> to NADPH is monitored at 340 nm using a microplate reader.
7. The increase in A<sub>340</sub> is used to calculate sugar content using a molar extinction coefficient (e) of 6,220 M<sup>-1</sup> cm<sup>-1</sup>.



*NB: To correct for the technical variation between microplate readers each value was corrected using a glucose standard curve measured on each individual plate.*

## 5.0 Appendix

### Stocks:



1. HEPES/KOH 100 mM, MgCl<sub>2</sub> 3 mM, pH 7.0 buffer [-20°C]
2. ATP 60mg/ml [-80°C]
3. NADP 36mg/ml [-80°C]
4. HK: Hexokinase 900 U•mL<sup>-1</sup> (*from Roche*: 1500•U mL<sup>-1</sup>, 120 µl suspension centrifuged, and pellet dissolved in 200 µl 0.1 M buffer) [-80°C]
5. PGI: Phosphoglucose isomerase 52 U•mL<sup>-1</sup> (*from Roche*: 175 U•mL<sup>-1</sup>, 60 µl suspension centrifuged, and pellet dissolved in 200 µl 0.1 M buffer) [-80°C]
6. INV: Invertase (*from Sigma*– lyophilized powder – dissolve as much as possible of the powder in 200 µl 0.1 M buffer)